



# ASAP report

**Reporter: Yi Xiao**

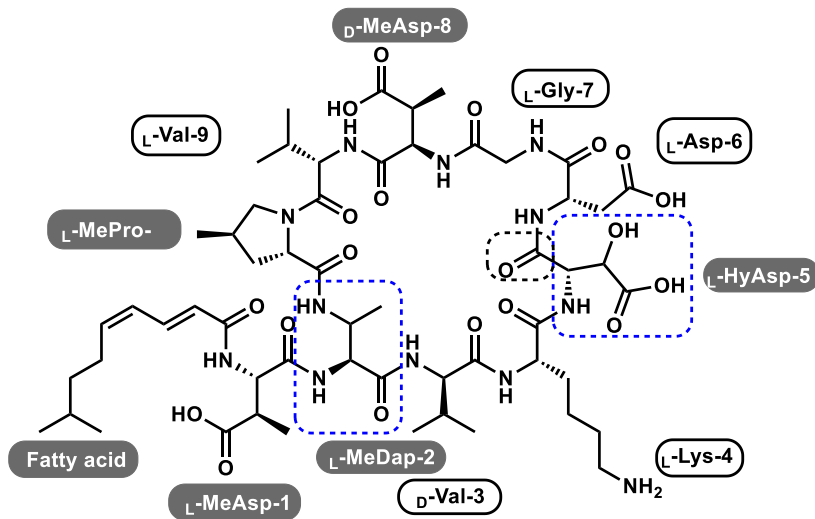
**Supervisors: *Prof. Tao Ye***

***Dr. Yian Guo***

***August 10th, 2020***

# Total Synthesis of Malacidin A by $\beta$ -Hydroxyaspartic Acid Ligation Mediated Cyclization and Absolute Structure Establishment

Zhenquan Sun<sup>[a]</sup>\*, Zhuo Shang<sup>[c]</sup>\*, Nicholas Forelli<sup>[c]</sup>, Kathy Hiu Lam Po<sup>[d]</sup>, Sheng Chen<sup>[d]</sup>, Sean F. Brady<sup>\*[c]</sup> and Xuechen Li<sup>\*[a, b]</sup>



Malacidin A

1a: L-(3S)-MeDap, L-(3R)-HyAsp

1b: L-(3R)-MeDap, L-(3R)-HyAsp

1c: L-(3S)-MeDap, L-(3S)-HyAsp

1d: L-(3R)-MeDap, L-(3S)-HyAsp



# Introduction

## Dr. Xuechen Li

Professor

Department of Chemistry &

State Key Laboratory of Synthetic Chemistry



2018- Professor, HKU

2014-2018 Associate Professor, HKU

2009-2014 Assistant Professor, HKU

2007-2009 Postdoc, Memorial Sloan Kettering Cancer Center  
(Professor Samuel Danishefsky )

2004-2007 Ph.D, Harvard University (Professor Dan Kahne)

2000-2003 M.Sc, University of Alberta (Professor Ole  
Hindsgaul)

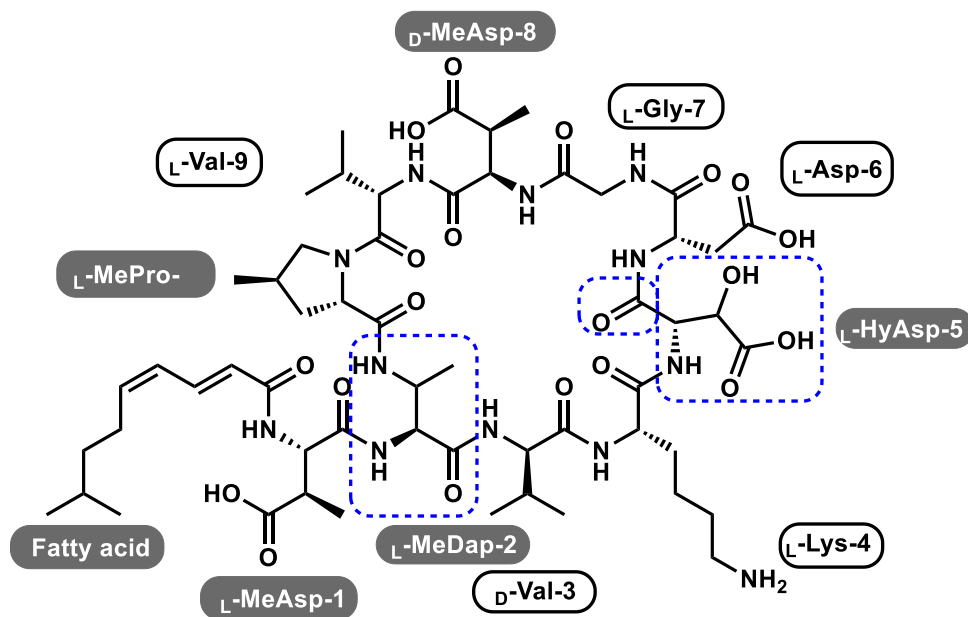
1995-1999 B.Sc, Nankai University

## **Research**

- I. Method developing for Bio-macromolecules synthesis
- II. Medicinal Chemistry of Natural Products
- III. Function Definition of Macromolecules and their PTMs
- IV. Drug Development



# Introduction



Malacidin A

- |                                |                                |
|--------------------------------|--------------------------------|
| 1a: L-(3S)-MeDap, L-(3R)-HyAsp | 1b: L-(3R)-MeDap, L-(3R)-HyAsp |
| 1c: L-(3S)-MeDap, L-(3S)-HyAsp | 1d: L-(3R)-MeDap, L-(3S)-HyAsp |

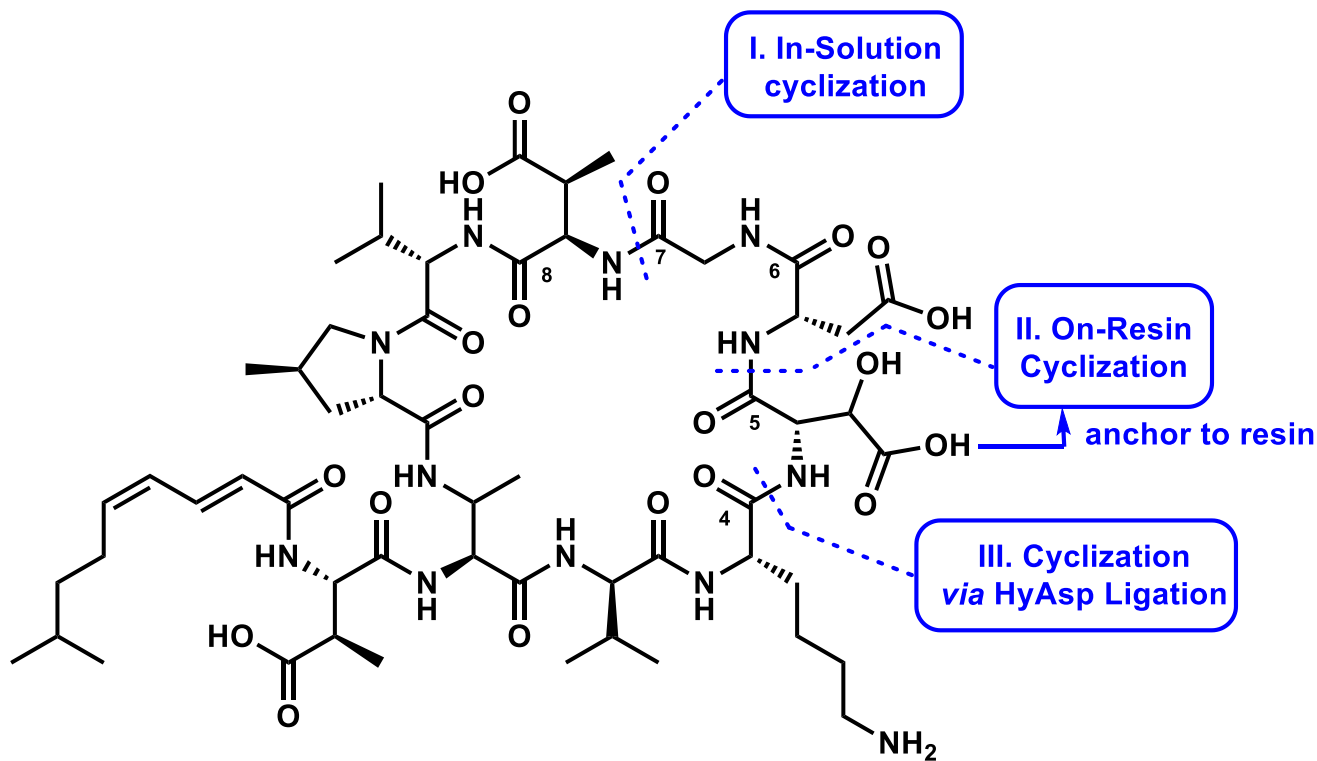
## Biology activity:

Malacidin A exhibits broad activity against Gram-positive bacteria including methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant *Enterococcus* (VRE), and has potent MIC values (0.2-2  $\mu\text{g/mL}$ ) in the presence of the divalent cation calcium.



# Total Synthesis of Malacidin A

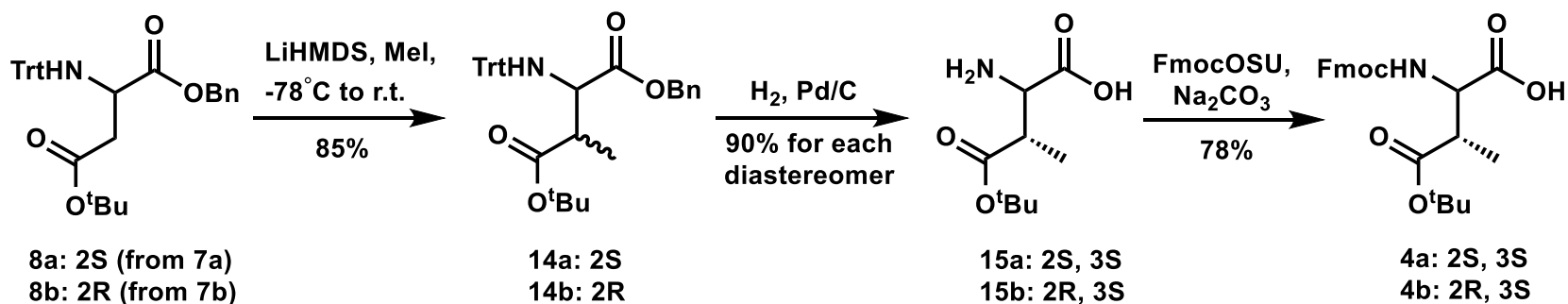
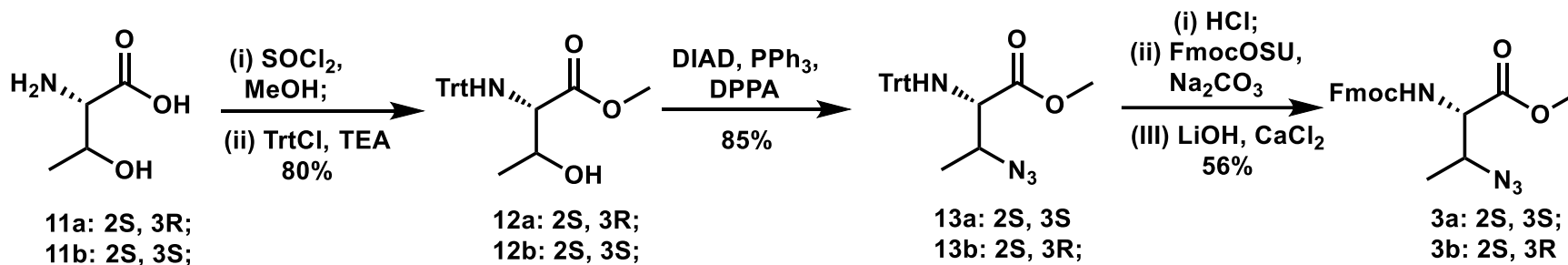
## Retrosynthesis





# Total Synthesis of Malacidin A

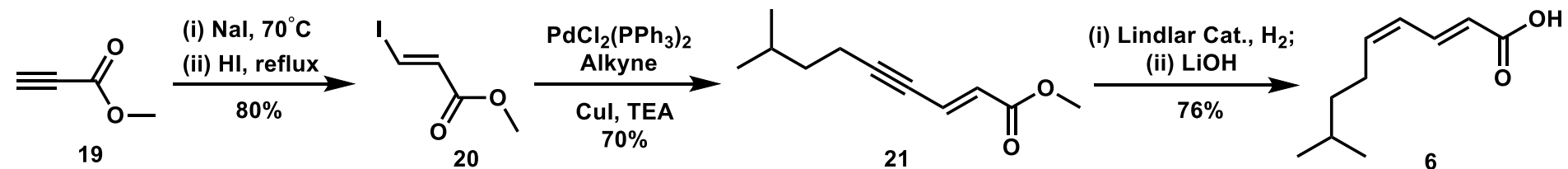
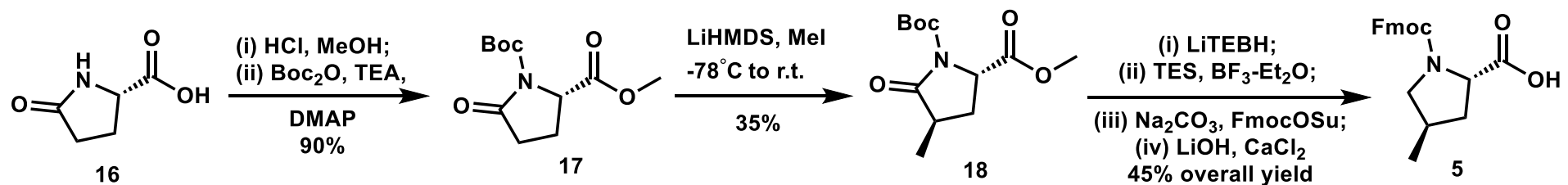
## Preparation of 3,4,5,6





# Total Synthesis of Malacidin A

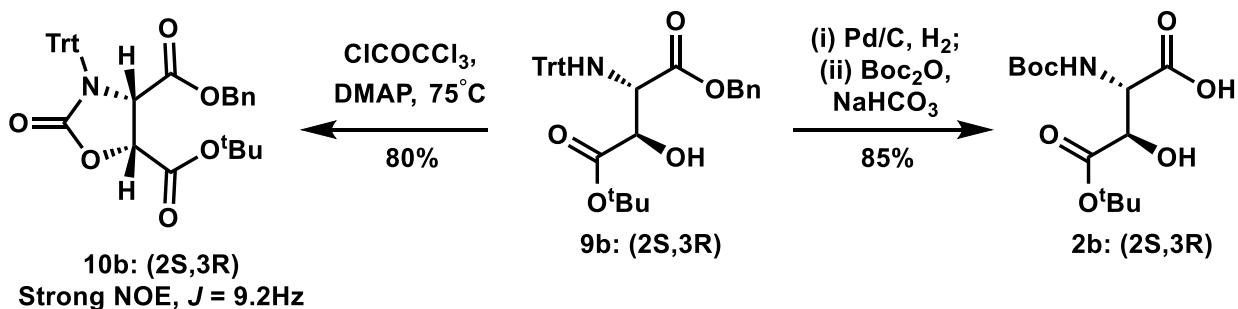
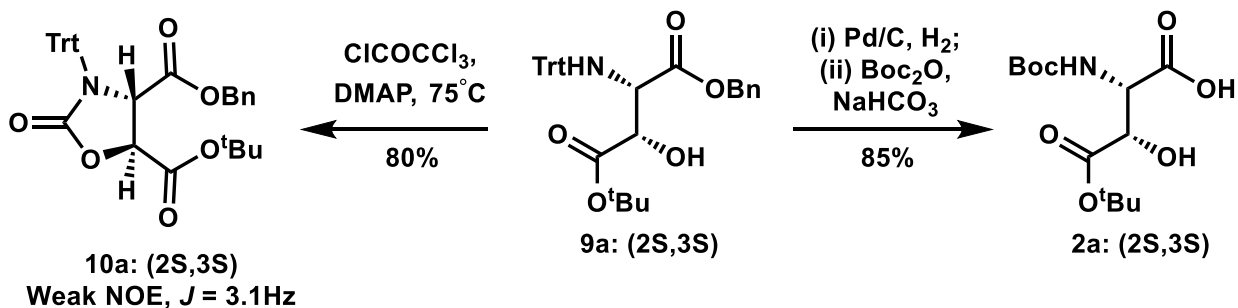
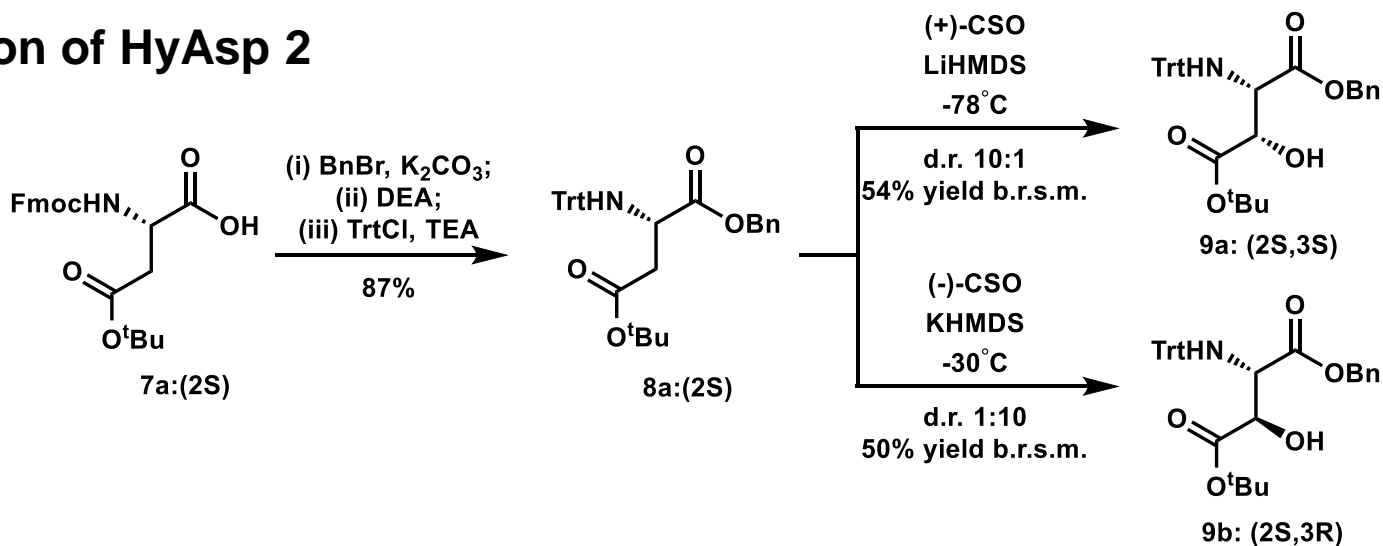
## Preparation of 3,4,5,6





# Total Synthesis of Malacidin A

## Preparation of HyAsp 2







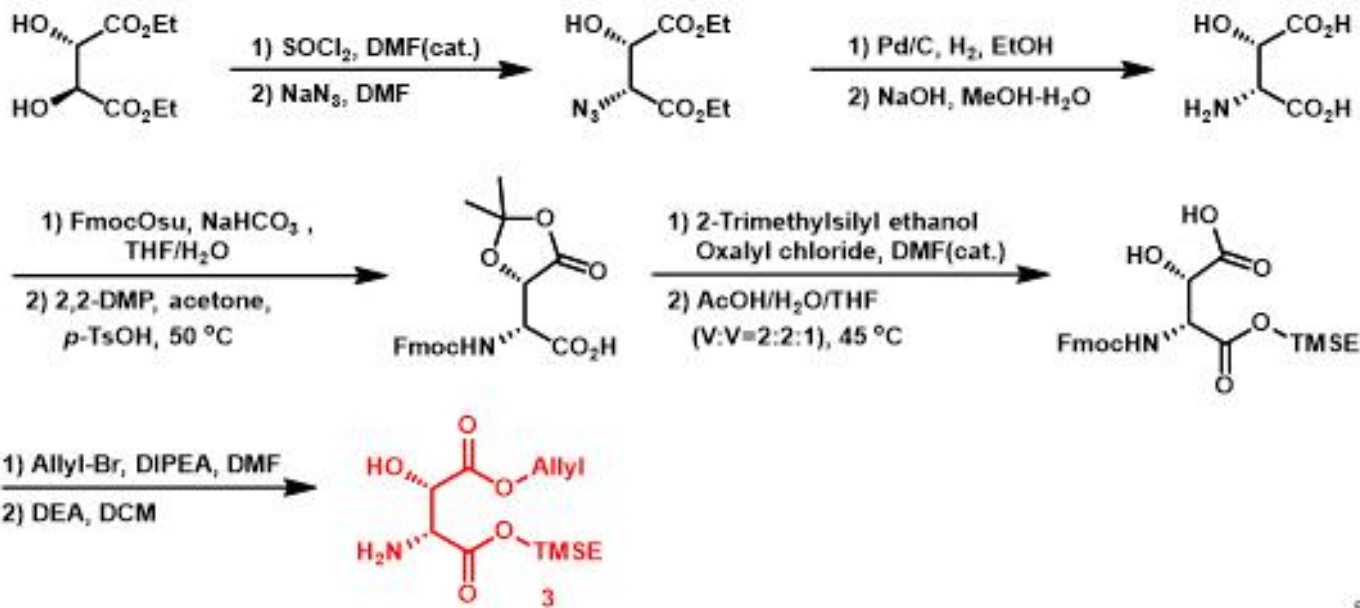
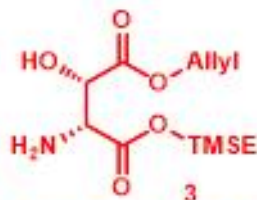
# Total Synthesis of Malacidin A

## Preparation of HyAsp 2

### Project Progress



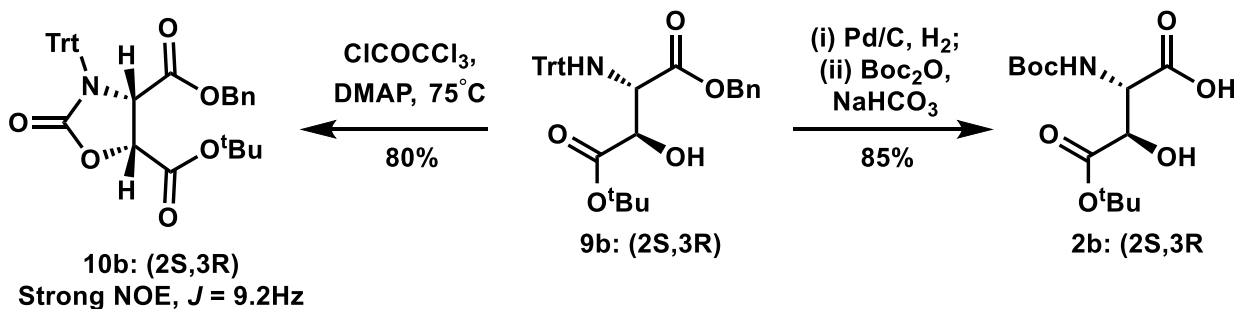
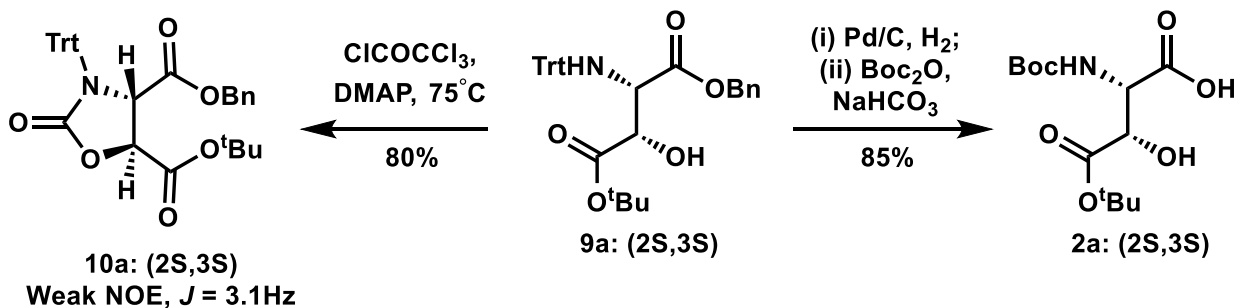
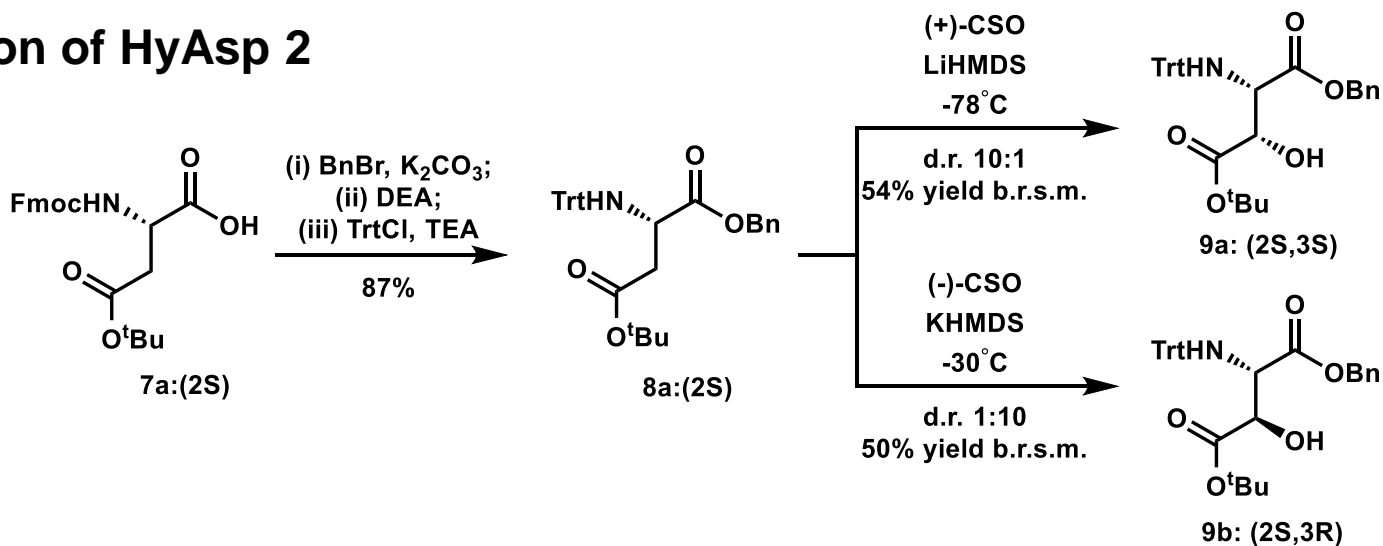
#### Synthesis of fragment 3





# Total Synthesis of Malacidin A

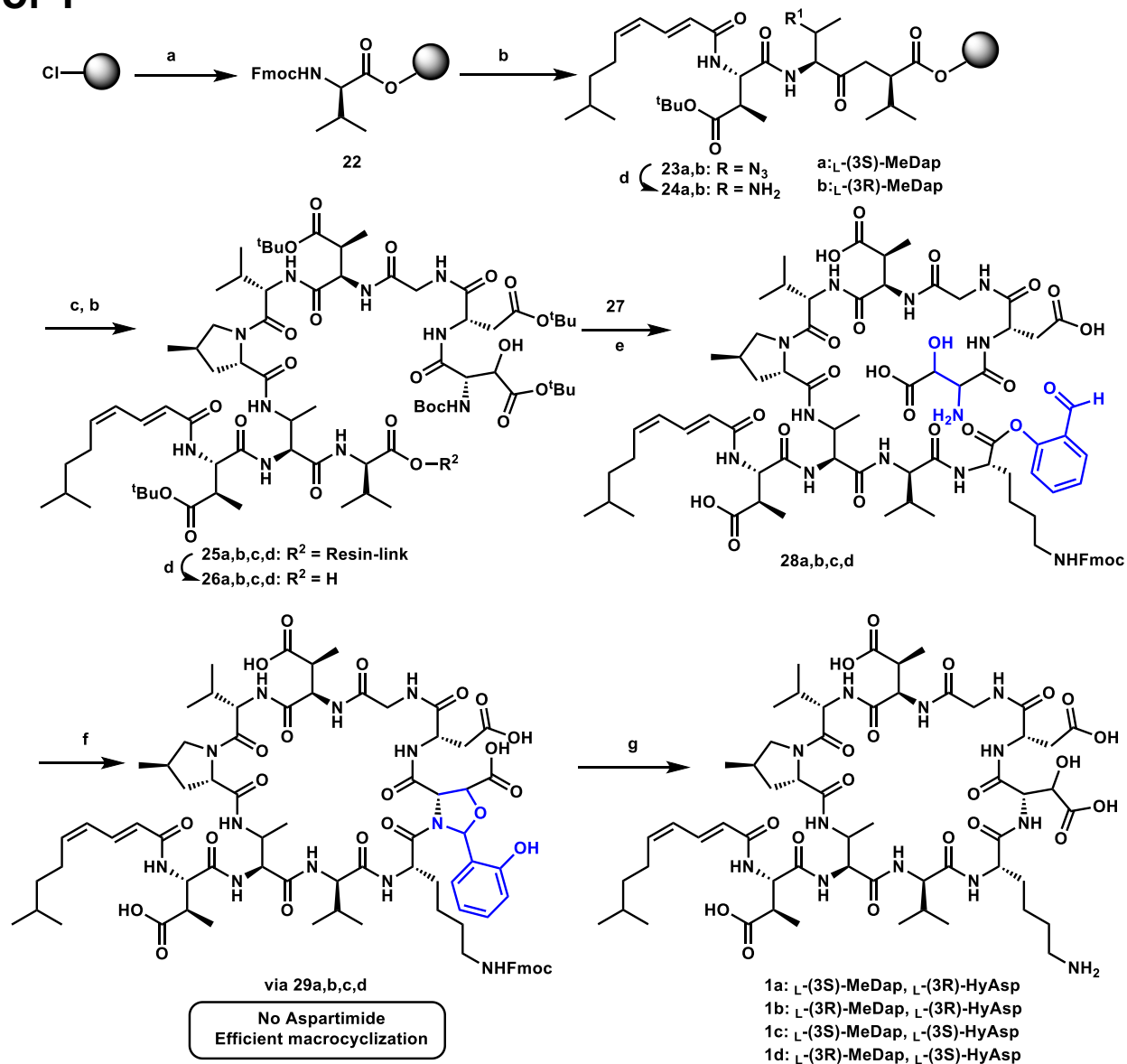
## Preparation of HyAsp 2





# Total Synthesis of Malacidin A

## Preparation of 1

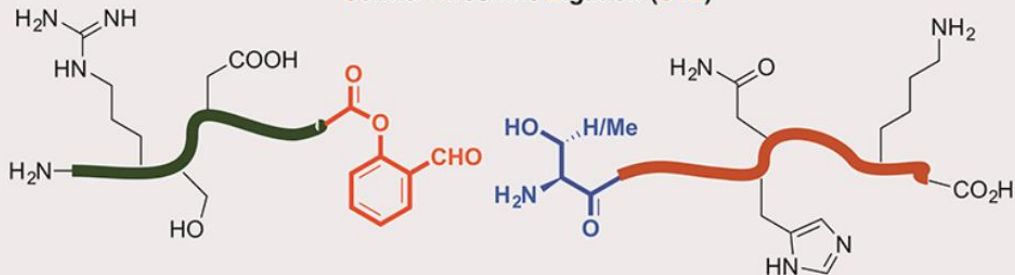




# Total Synthesis of Malacidin A

## Take-home message - Serine/Threonine Ligation

### Serine/Threonine Ligation (STL)

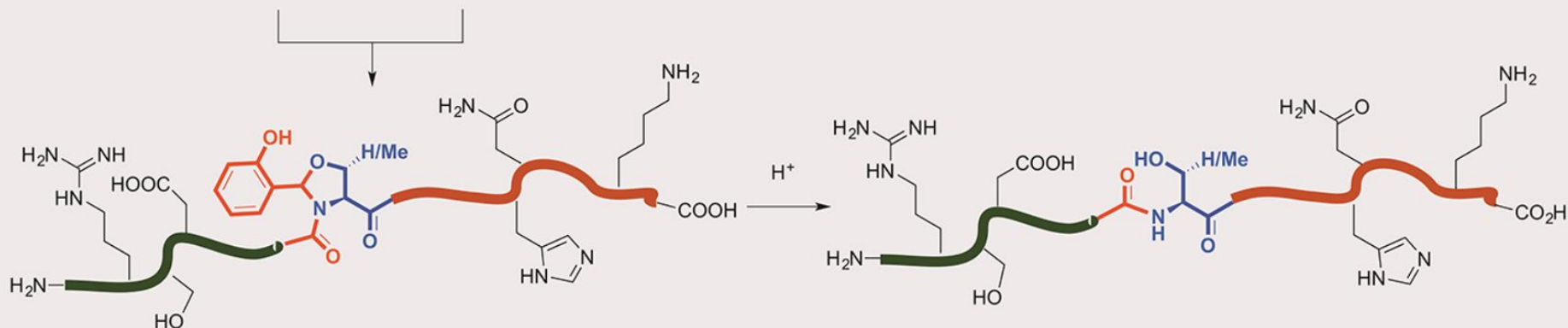


### Proteins

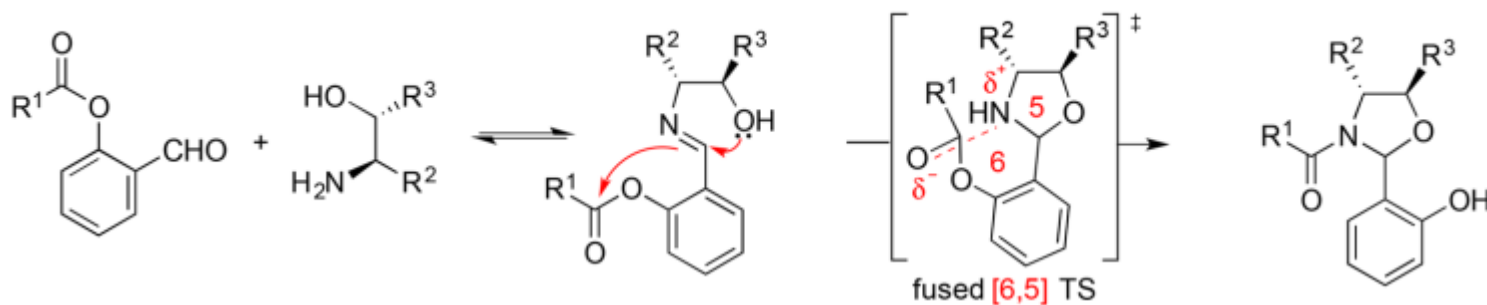
*ACYP* (2013)  
*MUC1* (2013)  
*IL 25* (2016)  
*HMGA1a* (2016)

### Cyclic peptides

*Daptomycin and analogues* (2013 & 2015)  
*Cyclomontanin B* (2013)  
*Yunnanin C and analogues* (2014)  
*Teixobactin and analogues* (2016-2018)



### c) [1,5] acyl transfer





## Acknowledgement

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**THANKS  
FOR YOUR ATTENTION!**

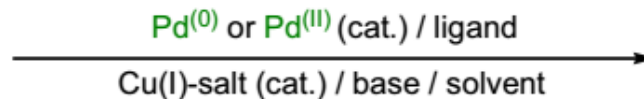


# Sonogashira cross-coupling

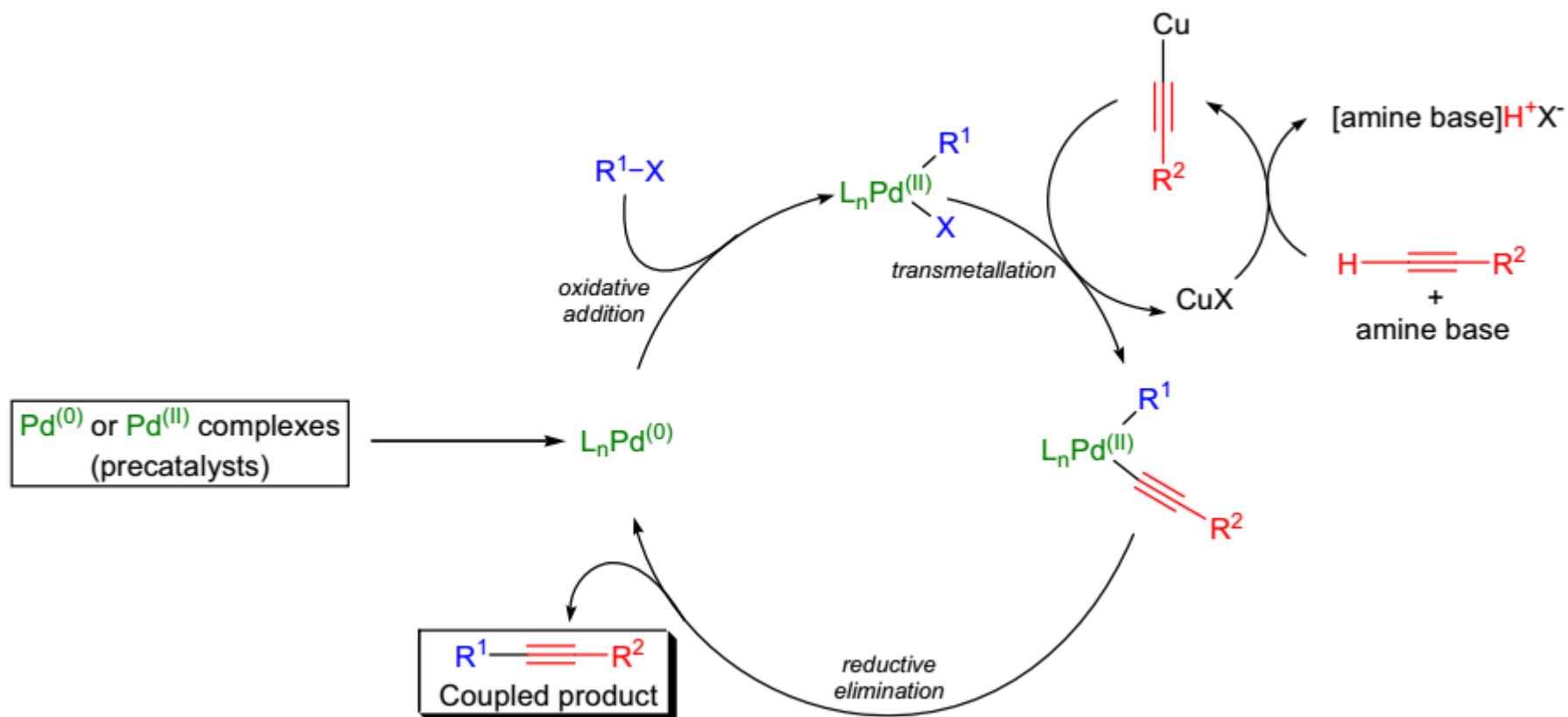
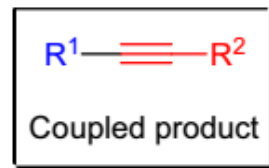
$R^1-X$   
 $R^1 = \text{aryl, alkenyl, heteroaryl}$   
 $X = \text{Cl, Br, I, OTf}$

+

$H-C\equiv C-R^2$   
 $R^2 = \text{H, alkyl, aryl, alkenyl, SiR}_3$



**Pd-catalyst:**  $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$  or  $\text{Pd}(\text{PPh}_3)_4$   
**Cu(I)-salt:**  $\text{CuI}$  or  $\text{CuBr}$   
**base:**  $\text{Et}_2\text{NH}$ ,  $\text{Et}_3\text{N}$ ,  $(\text{Chx})_2\text{NH}$ ,  $(i\text{-Pr})_2\text{NEt}$   
**solvent:**  $\text{MeCN}$ ,  $\text{THF}$ ,  $\text{EtOAc}$



### Scheme 1

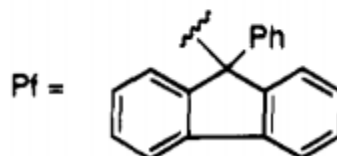
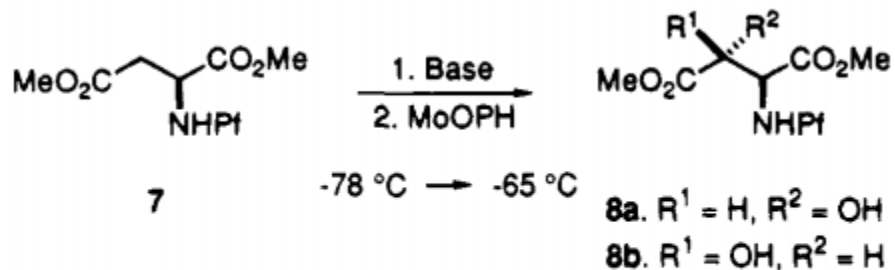
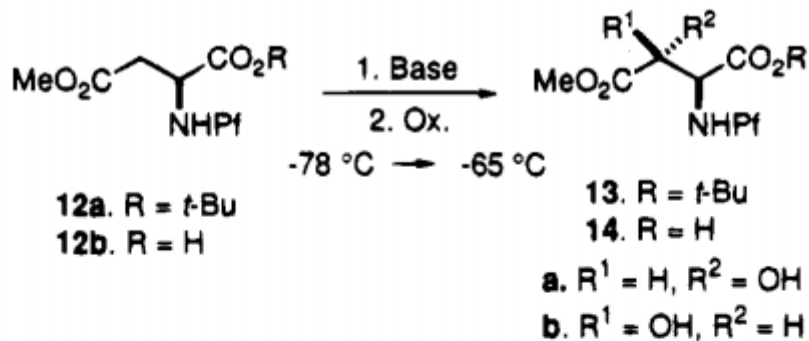


Table 1

entry	base (mol %)	cosolvent	8b/8a	yield (%)
1	KHMDS (180)	none	3/1	90 <sup>a</sup>
2	LHMDS (300)	none	1/8	65 <sup>a</sup>
3	LDA (300)	none	2/1	45 <sup>a</sup>
4	LTMP (180)	none	1/2	22 <sup>a</sup>
5	LHMDS (300)	DMPU	8/1	92 <sup>a</sup>
6	LHMDS (300)	HMPA	11/1	74 <sup>b</sup>
7	LHMDS (300)	DME	1/2.5	70 <sup>a</sup>
8	LHMDS (300)	TMEDA	1/8	80 <sup>a</sup>
9	LHMDS (300)	PMDET <sup>c</sup>	1/5	75 <sup>a</sup>
10	KHMDS (180)	18-crown-6	<i>d</i>	
11	LHMDS (300)	12-crown-4	<i>d</i>	
12	n-BuLi (100)/LHMDS (300)	none	1/20	60 (83) <sup>b</sup>
13	n-BuLi (100)/LHMDS (300)	HMPA	2/1	95 <sup>a</sup>

### Scheme 3



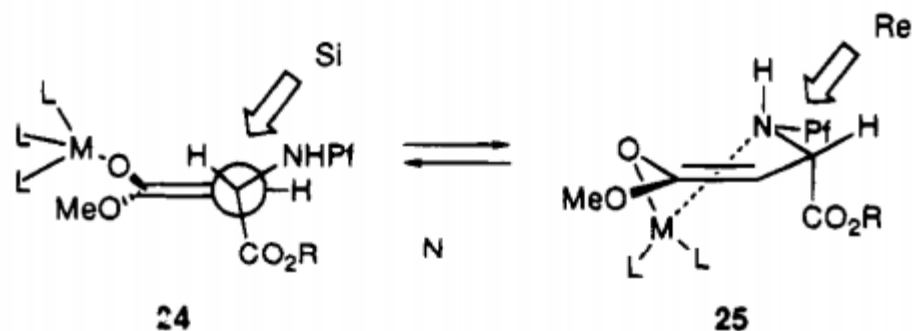
12a  $\rightarrow$  13

Base (cosolv.)	Oxidant	13b/13a	Yield <sup>a</sup>
KHMDS	MoOPH	7/1	95% (65%) <sup>b</sup>
LHMDS	MoOPH	-	-
LHMDS (HMPA)	MoOPH	4/1	70%
KHMDS	Davis' oxaz.	1/2	98%

12b  $\rightarrow$  14

Base (cosolv.)	Oxidant	14b/14a	Yield <sup>a</sup>
KHMDS	MoOPH	2/1	30%
LHMDS or LDA	MoOPH	-	-
LHMDS (HMPA)	MoOPH	-	-
KHMDS	Davis' oxaz.	7/1	86%
LHMDS	Davis' oxaz.	1/4	80%





Favored by good ligands of M  
(M = Li<sup>+</sup>) or M = K<sup>+</sup>

Favored by poor ligands of M  
and M = Li<sup>+</sup>

proach of an electrophile which cannot complex to the metal cation (MoOPH) to the less hindered face of the predominant nucleophilic species in the reaction medium should yield the observed results (Table 1, Scheme 3). The  $K_{eq}$  of the aforementioned equilibrium should be strongly dependent on the nature of the enolate counterion (M) and its ligands (L): the open form **24** would be favored by strongly coordinating ligands (DMPU or HMPA, Table 1, entries 5 and 6), or by the use of K<sup>+</sup> as the enolate counterion. Poorly coordinating ligands (HMDS or THF) and/or deprotonation of the Pf-amino group (by addition of 100 mol % of n-BuLi, entry 12) would favor the chelated form **25**.